- B light on cutaneous immune responses of humans with deeply pigmented skin. J Invest Dermatol 1991, 97, 729-744.
- Yarosh DB, Tsimis J, Yee V. Enhancement of DNA repair of UV damage in mouse and human skin by liposomes containing a DNA repair enzyme. J Soc Cosmet Chem 1990, 41, 85-92.
- Yarosh DB, Kibitel JT, Green LA, Spinowitz A. Enhanced unscheduled DNA synthesis in UV-irradiated human skin explants treated with T4N5 liposomes. J Invest Dermatol 1991, 97, 147-150.
- Kripke M, Cox P, Alas LG, Yarosh D. Pyrimidine dimers in DNA initiate systemic immunosuppression in UV-irradiated mice. Proc Natl Acad USA 1992, 89, 7516-7520.
- 10. Weinstock MA, Colditz GA, Willet WC, et al. Melanoma and the
- sun; the effects of swimsuits and a "healthy" tan on the risk of nonfamilial malignant melanoma in women. Am J Epidemiol 1991, 134, 462-470.
- de Gruijl FR, Sternborg HJCM, Forbes PD, et al. Wavelength dependence of skin cancer induction by ultraviolet irradiation of albino hairless mice. Cancer Res 1993, 60, 53-60
- Setlow RB, Grist E, Thompson K, Woodhead AD. Wavelengths effective in the induction of malignant melanoma. Proc Natl Acad Sci USA 1993, 90, 6666-6670

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V. Cutaneous Melanoma: Genetics and Molecular Biology

M. Pierotti

THE PURPOSE of genetic analysis of malignant melanoma is to identify genes involved in the transformation of melanocytes and melanoma tumour cell progression. Three basic approaches have been used to achieve this aim:

- (a) Genetic linkage analysis on familial melanoma to identify the chromosomal location of genes which predispose individuals to melanoma.
- (b) Cytogenetic analysis of tumour cells to identify frequently rearranged regions of the genome, where genes relevant to onset and/or progression of melanoma cells are located.
- (c) Molecular analysis of melanomas to identify mutated oncogenes or tumour suppressor genes playing crucial roles in melanoma development.

A brief summary of the present state of the art will follow for all the three above-reported approaches.

- (a) To date, the results of the studies of familial linkage analysis to identify patients who are at increased risk of developing melanoma are controversial. Bales et al. [1] reported that the gene for hereditary dysplastic nevus syndrome (HDNS) was on the distal part of the short arm of chromosome 1 (1p36). However, van Haeringen et al. [2] performed linkage studies in six large Dutch families with HDNS and did not find evidence of linkage between the putative loci for HDNS and chromosome 1p. The latter conclusion was also supported by a recent analysis of Lynch et al. [3].
- (b) Although non-random chromosomal rearrangements of chromosomes 1, 6 and 7 have been frequently reported in melanoma, consistent changes have also been detected for chromosomes 2, 3, 4, 10 and 11, and other genetic loci have been found to be affected in this tumour (for review see [4]). Interestingly, Lynch et al. [5] have recently completed cytogenetic studies with two kindreds affected by familial melanoma and have found evidence of chromosome instability that is dominantly inherited. Clonal cytogenetic abnormalities were also demonstrated in the skin and naevi of affected patients. The breakage abnormalities

- tended to involve chromosomes 14, 3, 1, 6, 11 and 22 (in order of decreasing frequency) [5]. Correlation of these cytogenetics results with linkage data may point to possible loci or frequently involved chromosomes containing a gene (or genes) responsible for FAMMM syndrome.
- (c) Oncogenes and growth factors. Early analysis by transfection of NIH/3T3 cells indicated the presence of the activated ras gene family in approximately 10% of the examined melanomas [6]. In addition, infection of melanocytes with retrovirus containing mutated ras genes resulted in a series of transformation-related changes [7], including abnormalities of chromosomes 6 (Albino A, Sozzi G, personal communication). Consequently, although it is now evident that activated ras genes are capable of conferring many of the characteristics of tumour progression on virus infected melanocytes, the low frequency of ras activation in melanoma in vivo suggests that alternative pathways from normal to transformed melanocytes still account for most human melanomas. In this context, a constitutive expression of the basic fibroblast growth factor (bFGF) gene has been reported in metastatic melanoma cell lines that are able to proliferate in the absence of added growth factor [8]. Although bFGF is not produced by normal melanocytes, the relevance of this finding waits to be assessed. Other oncogenes have been implicated in melanoma, mainly by their chromosomal localisation coincindent with recurrent abnormalities such as c-myb on chromosome 6g22 or EGF-R on chromosome 7 p12-p13. However, no firm association of these oncogenes with melanoma has ever been made. A gene which predisposes fish to melanoma and shows a high degree of homology to EGF-R has been identified in Xiphophorus [9]. This gene (Tu) represents a novel gene involved in the development of melanoma at least in that animal model.

Finally, recent indirect evidence points at ret, a tyrosine kinase receptor gene, as a gene potentially altered in melanoma. In fact, transgenic mice carrying the mouse metallothionein ret fusion gene were found to develop a

- severe melanosis and melanocytic tumours [10, 11]. Ret is found activated by rearrangement in a consistent number of papillary thyroid carcinomas [12], and, although not fully explored in melanomas, its expression has been detected in tumours originating from the neural crest [13, 14].
- (d) Tumour suppressor gene. Loss of heterozygosity (LOH) analysis of melanoma cells has indicated a high frequency of LOH at many loci on different chromosomes, again including chromosomes 1, 3, 6 and 9, but in general has also indicated a significant high chromosomal instability of these tumour cells (reviewed in [4]).

With another approach, using cell fusion techniques, Trent et al. [15] showed that the introduction of a normal chromosome 6 in two different human melanoma cell lines (one with a detectable 6q15 deletion) resulted in altered cell morphology and diminished cloning efficiency in soft agar, and the in vivo growth in nude mice correlated with the loss of the introduced chromosome 6. Unfortunately, no progress has been reported subsequently for the identification of the locus on chromosome 6 responsible for the suppression of malignant phenotype in melanoma. Finally, two recent papers have reported a very high frequency of positivity (85%) by immunostaining with p53 antibodies in two series of 83 specimens of primary and metastatic melanomas.

These findings represent one of the highest incidences of p53 mutation yet registered in a human malignancy and support the concept that alterations of this gene may be an early event in melanoma development [16, 17]. So far, a similar analysis also employing molecular techniques has not been reported.

In conclusion, although in recent years several investigations have dealt with the issue of genetics and molecular biology of melanomas, perhaps with the exception of p53, we are still far from the identification of relevant genes significantly involved in its development and progression. However, the success of the molecular approach in identifying the genes of other inherited cancer syndromes, such as retinoblastoma or adenomatous polyposis (for review see [18]), leaves us with the hope that future investigations on the molecular aspect of melanoma will provide the clue for a more successful management of this increasingly important neoplastic disease.

- Bales SJ, Dracopoli NC, Tucker MA, et al. Mapping the gene for hereditary cutaneous malignant melanoma dysplastic nevus to chromosome 1p. N Eigl J Med 1989, 320, 1367-1372.
- Van Haeringen A, Bergman W, Nelen MR, et al. Exclusion of the dysplastic nevus syndrome (DNS) locus from the short arm of chromosome 1 by linkage studies in Dutch families. Genomics 1989, 5, 61-64.
- Lynch HT, Fusaro RM. The surgeon, genetics and malignant melanoma. Arch Surg 1992, 127, 317-320.
- Fountain JW, Bale SJ, Housman DE, Dracopoli NC. Genetics of melanoma. Cancer Surveys 1990, 9, 645-672.
- Lynch MT, Furaro RM, Sandberg AA, Bixenman H. Chromosome breakage in the FAMMM syndrome. Cancer, in press.
- Albino AP, Le Strange R, Oliff AI, Furth ME, Old LJ. Transforming ras genes from human melanoma: a manifestation of tumor heterogeneity? Nature 1984, 308, 69-72.
- Albino AP, Houghton AN, Eisinger M, et al. Class II histocompatibility antigen expression in human melanocytes transformed by Harvey murine sarcoma virus (Ha-MSV) and Kirsten MSV retroviruses. J Exp Med 1986, 164, 1710-1722.
- Halaban R, Kwon BS, Ghosh S, Delli-Bovi P, Baird A. bFGF is an autocrine growth factor for human melanomas. Oncogene Res 1988, 3, 177-186.
- Wittbrodt J, Adam D, Malitschek B, et al. Novel putative receptor tyrosine kinase encoded by the melanoma inducing Tu locus in Xiphophorus. Nature 1989, 341, 415-421.
- Iwamoto T, Takahashi M, Ito M, et al. Aberrant melanogenesis and melanocytic tumour development in transgenic mice that carry a metallothionein/set fusion gene. EMBO 7 1991,10, 3167-3176.
- Iwamoto T, Takahashi M, Ohbayashi M, Nakashima I. The ret oncogene can induce melanogenesis and melanocyte development in Wv/Wv mice. Exp Cell Res 1992, 200, 410-415.
- Pierotti MA, Santoro M, Jenkins RB, et al. Characterization of an inversion on the long arm of chromosome 10 juxtaposing D10S170 and RET and creating the oncogenic sequence RET/PTC. Proc Natl Acad Sci USA 1992, 89, 1616-1620.
- 13. Santoro M, Rosati R, Grieco M, et al. Oncogene 1990, 5, 1595-1598.
- Ikeda I, Ishizaka Y, Tahira T, et al. Specific expression of the ret proto-oncogene in human neuroblastoma cell lines. Oncogene 1990, 5, 1291–1296.
- Trent JM, Stanbridge EJ, McBride HL, et al. Tumorigenicity in human melanoma cell lines controlled by introduction of human chromosome 6. Science 1990, 247, 568-571.
- Stretch JR, Gatter KC, Ralfkiaer E, Lane DP, Harris AL. Expression of mutant p53 in melanoma. Cancer Res 1991, 51, 5976-5979.
- Aksen LA, Morkve O. Expression of p53 protein in cutaneous melanoma. Int J Cancer 1992, 52, 13-16.
- Pierotti MA, Dragani TA. Genetics and cancer. Curr Opinion Oncol 1992, 4, 127–133.

VI. What Has Been Achieved by Primary and Secondary Prevention Campaigns?

R. Mackie

THE THREE main types of skin cancer are basal cell cancer, squamous cell carcinoma and malignant melanoma. In the past, basal and squamous cell carcinomas have tended to be grouped together as non-melanoma skin cancer, but there are good reasons for separating these two entities out. The exact pattern of incidence in relation to sun exposure is different between the

two malignancies, and it is becoming clear that squamous cell carcinoma is a greater risk in those who are immunosuppressed.

A further problem that arises regarding basal cell and squamous cell carcinomas is incomplete cancer registration. This is because there is a continuing tendency to diagnose a proportion of these lesions clinically, and to then treat with non-excisional